

The Synthesis of Tricyclic Cocaine Analogs via the 1,3-Dipolar Cycloaddition of Oxidopyridinium Betaines.

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Abstract: Tricyclic cocaine analogs with a spatially fixed nitrogen lone pair were synthesized as structural probes of the dopamine transporter. A tandem cycloaddition/radical cyclization protocol was used to gain access to analog 12 with a two carbon linker between C-2 and N-8. On the other hand, the intramolecular dipolar cycloaddition reaction of betaine 13 was used to procure cocaine analog 18 and its C-3 epimer 19 in which an extra ring links N-8 to C-6. Binding studies reveal 12c and 18 to be potent DAT ligands. © 1997 Elsevier Science Ltd. All rights reserved.

Cocaine (1) has long been recognized to act as a potent central nervous system stimulant. Addicts may lose the ability to function at work or in interpersonal interactions. As a result, cocaine is one of the greatest concerns of the American public today. It is clear that immediate therapies are needed for the treatment of individuals who have become addicted to this powerful reinforcing drug. A growing body of evidence points to the ability of cocaine to bind to the dopamine transporter (DAT) and to inhibit the reuptake of dopamine (DA) as being responsible for the reinforcing properties of this drug. A number of highly potent cocaine analogs together with information concerning their structure-activity relationships (SAR) at the DAT have been reported. However, the precise details of the binding interactions between these analogs and the DAT is still a matter of much discussion.

Previously it has been shown that replacement of the C-3 benzoate by phenyl leads to higher potency cocaine analogs (2; these phenyl bearing structures are often referred to as the WIN series). Additionally, we have shown that cocaine's C-2 ester group can be replaced by alkyl and alkenyl groups as in 3 and still retain high binding affinity. A number of N-modified cocaine analogs have been reported. For example, replacement of the methyl group of cocaine by electronically similar but sterically larger groups has marginal effect on binding activity. Replacement of the 8-methyl group by propyl has almost no effect, while replacement by allyl or benzyl reduces activity by a factor of 7-fold or less. N-substitution with electron-withdrawing groups can significantly decrease activity, however, a basic amine or nitrogen is not always required for potent activity. Little is known experimentally about the spatial requirements of the nitrogen lone pair of these

cocaine analogs. The directionality of the nitrogen lone pair is, however, likely to be of some consequence to binding affinity. Conformational analysis of a series of methylphenidate derivatives and their N-methyl analogs showed that the reduced binding affinities of the latter could be explained by the spatial requirements at nitrogen. The added N-methyl group was shown to preferentially occupy the space presumed to be preferred by the lone pair of cocaine. Semi-empirical quantum chemistry (PM3) calculations along with mutational data suggest that cocaine binds in its neutral form to the dopamine transporter forming a weak hydrogen bond involving a protonated aspartate (Asp 79) residue from the dopamine transporter. Herein we report the synthesis of conformationally restricted cocaine analogs that may be used to further map the cocaine/receptor interactions.

The synthetic route employed in the present study (Scheme 1) exploited the versatile 1,3-dipolar cycloaddition of 3-hydroxypyridinium betaines with electron deficient olefins for the synthesis of the tropane skeleton. This synthesis commences with the preparation of the 4-(p-substituted)phenylpyridines 5b and c (X = Cl, Me) by the route previously reported for the unsubstituted (X = H) analog 5a. N-Alkylation with 2,3-dibromopropene in THF afforded the pyridinium bromide salts 6b and 6c in 95% yield. The tandem cycloaddition/radical cyclization methodology reported by Ghosh and Hart was used to produce the desired tricyclic ketones 8b and 8c. Thus, the dipolar cycloaddition of the betaine of 6b or 6c (generated in situ with Et_3N) with phenyl vinyl sulfone afforded a mixture of the 6- and 7-exo-phenylsulfonyl regiosomers 7b or 7c (in 60 and 21% isolated yield, respectively, from 6). Radical cyclization of 7b afforded the tricyclic ketone 8b as a white crystalline solid in 54% yield (75% for the analogous transformation of 7c into 8c). The structure of 8b was unequivocally confirmed by X-ray analysis.

Scheme 1. Reagents and conditions: (a) CuI, LiCl, PhOC(O)Cl, Et₂O then p-XPhMgBr; (b) o-chloranil, toluene; (c) 20% Pd(OH)₂/C, H₂, MeOH; (d) 2,3-dibromopropene, THF; (e) phenyl vinyl sulfone, Et₃N, MeCN, reflux; (f) n-Bu₃SnH, AIBN, toluene; (g) DIBAL, CH₂Cl₂; (h) 6% Na(Hg), Na₂HPO₄, MeOH; (i) n-BuLi, THF then PhOC(S)Cl.

Reduction of the ketone **8b** or **8c** at -78 °C afforded a single isomer in good yield (84 and 79%, respectively). Reductive desulfonylation of **9b** resulted in the partial reduction of the aromatic chloride to afford an inseparable mixture of **10a** and **10b**. The

reductive desulfonylation of 9c readily afforded alcohol 10c in 63% yield. Barton deoxygenation then afforded the desired cocaine analog 12c (in 16% yield for 2 steps). In this rigid analog, the direction of the nitrogen lone pair is fixed so as to point toward the two carbon bridge of the tropane skeleton. The tether also provides an olefinic C-2 substituent. The subnanomolar affinity of the related 2β -vinyl- 3β -(p-chlorophenyl) analog of cocaine has been reported. 10

The tricylic cocaine analog containing a tether to the two carbon bridge of the tropane skeleton was prepared, exploiting the intramolecular dipolar cycloaddition¹⁴ of betaine 13. Compound 13 was readily prepared in 98% yield from pyridine 5c. The intramolecular cycloaddition was regioselective to afford the 6-bridged tropenone isomer 14 as the only cycloadduct detected (isolated in 36% yield). The structure of 14 was again confirmed by X-ray analysis. Luche reduction of the enone afforded a mixture of alcohols that were directly acylated to afford the epimeric allyl acetates 15. These acetates could be separated, or more conveniently the mixture (ca. 3:1 by NMR) was directly utilized in the CuCN catalyzed cross coupling as previously reported.⁹ This cross coupling afforded a complex mixture in which all four possible isomeric products could be detected (28:11:1:1, by GC/MS).¹⁵ Two major cross coupling products were isolated and tentatively assigned as 16 and 17 (42 and 20% yield), respectively, arising from β-addition of the butyl group. The reduction of the double bond of 16 under acidic conditions afforded a separable mixture of the cocaine analogs 18 and 19 (3:1 by GC/MS). The structure of 19 was confirmed by conversion to the p-toluenesulfonate salt and X-ray analysis.

5c
$$a, b$$
 $Ph-p-(Me)$ $O o o$ $O o$ O

Scheme 2. Reagents and conditions: (a) 4-bromo-1-butene, THF; (b) Amberlite IRA-400 (OH), MeOH; (c) xylenes, reflux; (d) CeCl₃•7H₂O, NaBH₄, MeOH; (e) Ac₂O, pyridine; (f) CuCN, n-BuMgBr, ether; (g) p-TsOH, 10% Pd/C, H₂, EtOAc.

In conclusion, synthetic pathways for the construction of cocaine analogs whose nitrogen lone pairs are spatially defined have been delineated. Preliminary biological experiments reveal that 12c and 18 exhibit substantial affinity for the DAT, with a binding affinity (K_i) of 25 ± 3 nM for 12c and 17 ± 1 nM for 18 as compared to 280 ± 60 nM for (-)-cocaine. These data would suggest that the directionality of the nitrogen lone pair is not a crucial determinant of high affinity binding. Studies to further address this issue are currently underway, and full details of this work will be reported separately.

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References and Notes

- 1. Lounasmaa, M. Alkaloids 1988, 33, 1-81.
- 2. (a) Woolverton, W. L.; Johnson, K. M. Trends Pharmacol. Sci. 1992, 13, 193-200; (b) Woolverton, W. L.; Kleven, M. S. NIDA Research Monograph 1988, 88, 160-184.
- 3. Volkow, N. D.; Wang, G. J.; Fowler, J. S.; Logan, J.; Gately, S. J.; Hitzemann, R.; Chen, A. D.; Dewey, S. L.; Pappas, N. *Nature* 1997, 6627, 830-833.
- 4. Clarke, R. L.; Daum, S. J.; Gambino, A. J.; Aceto, M. D.; Pearl, J.; Levitt, M.; Cumiskey, W. R.; Bogado, E. F. J. Med. Chem. 1973, 16, 1260-1267.
- 5. (a) Carroll, F. I.; Gao, Y.; Rahman, M. A.; Abraham, P.; Parham, K.; Lewin, A. H.; Boja, J. W.; Kuhar, M. J. J. Med. Chem. 1991, 34, 2719-2725; (b) Carroll, F. I.; Lewin, A. H.; Boja, J. W.; Kuhar, M. J. In Drug Design for Neuroscience; Kozikowski, A. P., Ed.; Raven Press: New York, 1993, pp. 149-166.
- 6. Carrol, F. I.; Lewin, A. H.; Boja, J. W.; Kuhar, M. J. J. Med. Chem. 1992, 35, 969-981.
- 7. Kozikowski, A. P.; Saiah, M. K. E.; Johnson, K. M.; Bergmann, J. S. J. Med. Chem. 1995, 38, 3086-3093.
- 8. (a) Abraham, P.; Pitner, J. B.; Lewin, A. H.; Kuhar, M. J.; Carrol, F. I. *J. Med. Chem.* 1992, 35, 141-144. (b) Stoelwinder, J.; Roberti, M.; Kozikowski, A. P.; Johnson, K. M.; Bergmann, J. S. *Bioorg. Med. Chem. Lett.*, 1994, 4, 303-308.
- 9. (a) Kozikowski, A. P.; Saiah, M. K. E.; Bergmann, J. S.; Johnson, K. M. J. Med. Chem. 1994, 37, 3440-3442. (b) Madras, B. K.; Pristupta, Z. P.; Nizik, H. B.; Liang, A. Y.; Blundell, P.; Gonzalez, P. C. Synapse 1996, 24, 340-346.
- 10. (a) Froimowitz, M.; Deutsch, H. M.; Shi, Q.; Wu, K.-M.; Glaser, R.; Adin, I.; George, C.; Schweri, M. M. Bioorg. Med. Chem. Lett. 1997, 7, 1213-1218. (b) Kozikowski, A. P.; Wang, S.; Simoni, D.; Roberti, M.; Manferdini, M.; Du, P.; Johnson, K. M. Submitted to J. Med. Chem. (c) Kitayama, S.; Shimada, S.; Xu, H.; Markham, L.; Donovan, D. H.; Uhl, G. R. Proc. Natl. Acad. Sci. USA 1992, 89, 7782-7785.
- 11. Dennis, N.; Katritzky, A. R.; Takeuchi, Y. Angew. Chem., Int. Ed. Engl. 1976, 15, 1-9.
- 12. Araldi, G. L.; Kozikowski, A. P.; Ball, R. B. J. Org. Chem. 1997, 62, 503-509.
- 13. All compounds described were purified by chromatography on silica gel, and were characterized by NMR and mass spectra. Spectroscopic data for selected compounds: 12c: ^{1}H NMR (CDCl₃) δ 1.39 (dd, 1 H), 1.77 (m, 1 H), 2.01-2.23 (m, 5 H), 2.34 (s, 3 H), 2.47 (s, 1 H), 3.26 (m, 1 H), 3.45 (m, 1 H), 3.35 (m, 2 H), 3.61 (s, 2 H), 4.25 (s, 1 H), 4.78 (s, 1 H), 7.07 (d, 2 H), 7.12 (d, 2 H). 18: (p-toluenesulfonate salt) ^{1}H NMR (CD₃OD) δ 0.75 (t, 3 H), 1.04-1.16 (m, 3 H), 1.25-1.34 (m, 3 H), 1.70-1.84 (m, 3 H), 2.02-2.16 (m, 4 H), 2.29 (s, 3 H), 2.36 (s, 3 H), 2.53 (m, 1 H), 2.67 (s, 1 H), 3.24 (m, 1 H), 3.47 (m, 1 H), 3.70 (m, 1 H), 3.83 (d, 1 H), 7.11 (s, 4 H), 7.25 (d, 2 H), 7.74 (d, 2H). 19: (p-toluenesulfonate salt) ^{1}H NMR (CDCl₃) δ 0.71 (t, 3 H), 0.74-1.00 (m, 3 H), 1.09-1.15 (m, 2 H), 1.57-1.83 (m, 4 H), 1.91 (m, 1 H), 2.07 (m, 1 H), 2.32 (s, 3 H), 2.37 (s, 3 H), 2.40 (m, 1 H), 2.80 (m, 2 H), 2.95 (m, 1 H), 3.17 (m, 1 H), 3.34 (ddd, 1 H), 3.81 (d, 1 H), 3.89 (s, 1 H), 4.09 (dd, 1 H), 6.95 (d, 2 H), 7.09 (d, 2 H), 7.14 (d, 2 H), 7.82 (d, 2 H).
- 14. Sammes, P. G.; Watt, R. A. J. Chem. Soc., Chem. Comm. 1976, 367-368.
- 15. The same isomeric mixture (by GC/MS) was also obtained using either of the isolated allyl acetates.
- 16. The K_i values were determined by displacement of [³H]mazindol binding. We thank Dr. Kenneth M. Johnson for providing the binding data.